

ENZYMATIC SYNTHESIS OF  $\gamma$ -AMINOBUTYRIC ACID USING  
IMMOBILIZED L-GLUTAMATE DECARBOXYLASE

R. P. Yanushyavichyute, A. B. Paulyukonis,  
and D. A. Kazlauskas

UDC 547.466.3.07

$\gamma$ -Aminobutyric acid (GAMA) is an important metabolite of nerve tissues and is widely used in medicine. In industry, GAMA is obtained by the hydrolysis of pyrrolidone [1]. Considerable interest is presented by a biotechnological method of obtaining GAMA by the  $\alpha$ -decarboxylation of L-glutamic acid. The production of GAMA by this method has been described previously with the use as decarboxylation biocatalyst of the water-soluble enzyme L-glutamate decarboxylase (GDC) or bacterial cells [2-6]. We have investigated the process of obtaining GAMA through the decarboxylation of L-glutamic acid by immobilized GDC.

The production and main properties of immobilized GDC have been described elsewhere. The enzyme was immobilized by the addition of a partially purified preparation of GDC from *Escherichia coli* to Silochrome CX-2,5 modified by the introduction of bromoacetamide groups. Decarboxylation was carried out in a thermostated (37°C) laboratory minireactor connected with a pH-stat apparatus. The reactor was charged with 50 mg of immobilized GDC having an activity of 400 U/g (U —  $\mu$ mole/min) and 25 ml of a 0.05 M solution of L-glutamic acid in which pyridoxal phosphate had previously been dissolved ( $5 \cdot 10^{-5}$  M) and the pH has been brought to a 4.6 with the aid of 0.1 N NaOH. On decarboxylation, the equivalent amount of OH<sup>-</sup> ions is liberated. A constant pH of 4.6 was maintained by titration with 0.2 N HCl with the aid of the pH-stat. By simultaneously recording the consumption of titrant it was possible to follow the degree of conversion and the yield of product. After decarboxylation of the first portion of the L-glutamate solution, a minipump was switched on and the solution of the substrate was fed at a constant rate of  $10^{-5}$  mole/min with the simultaneous removal of part of the reaction mixture in such a way that the total volume remained constant. Under these conditions a constant rate of decarboxylation of  $9.5 \cdot 10^{-6}$  mole/min was maintained, which corresponds to a 95% conversion of the substrate. The solution of the product was concentrated in vacuum and the GAMA was precipitated with ethanol. After reprecipitation from water with ethanol, a crystalline product was obtained with mp 200-201°C, which corresponds to the literature figure. Yield 85%. Thin-layer chromatography on Silufol plates on the ethanol-water (7:3) solvent system showed the identity of the GAMA obtained with a known sample ( $R_f$  0.38) and the absence of contamination by L-glutamic acid ( $R_f$  0.66).

The immobilized GDC can be used as biocatalyst repeatedly and the synthesis can be performed continuously. The process takes place at a low concentration of L-glutamic acid, which permits the inhibition by the substrate that is characteristic for GDC to be avoided.

LITERATURE CITED

1. J. E. Thompson, J. K. Pollard, and F. C. Steward, Arch. Biochem. Biophys., 46, 248 (1953).
2. M. N. Camien, L. E. Clure, A. Lepp, and M. S. Dunn, Arch. Biochem. Biophys., 43, 378 (1953).
3. K. Nakayama and H. Hagino, Japanese Patent No. 6901196 (1969); Chem. Abstr., 70, 86,262 (1969).
4. K. Nakayama and H. Tanaka, Japanese Patent No. 7015434; Chem. Abstr., 73, 65,021 (1970).
5. E. M. Gubarev and Yu. V. Galaev, Biokhimiya, 25, 261 (1960).
6. Yu. V. Galaev and E. G. Chuplygina, Abstracts of Lectures at an All-Union Conference on Amino Acids for Agriculture, the Foodstuffs Industry, Public Health, and Scientific Investigations [in Russian], Frunze (1981), p. 31.
7. R. P. Yanushyavichyute, A. B. Paulyukonis, and D. A. Kazlauskas, Prikl. Biokhim. Mikrobiol., 18, 357 (1982).

---

All-Union Scientific-Research Institute of Applied Enzymology, Vilnius. Translated from Khimiya Prirodnikh Soedinenii, No. 2, pp. 246-247, March-April, 1983. Original article submitted November 29, 1982.